

## TEMPORAL TRENDS OF PERFLUOROALKYL SUBSTANCES (PFAS) IN EGGS OF COASTAL AND OFFSHORE BIRDS: INCREASING PFAS LEVELS ASSOCIATED WITH OFFSHORE BIRD SPECIES BREEDING ON THE PACIFIC COAST OF CANADA AND WINTERING NEAR ASIA

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**Abstract:** Perfluoroalkyl substances (PFAS) such as perfluoroalkyl carboxylates (PFCAs) and perfluoroalkyl sulfonates (PFSAs) have become virtually ubiquitous throughout the environment, and, based on laboratory studies, have known toxicological consequences. Various national and international voluntary phase-outs and restrictions on these compounds have been implemented over the last 10 to 15 years. In the present study, we examine trends (1990/1991–2010/2011) in aquatic birds (ancient murrelet, *Synthliboramphus antiquus* [2009 only]; Leach's storm-petrels, *Oceanodroma leucorhoa*; rhinoceros auklets, *Cerorhinca monocerata*; double-crested cormorants, *Phalacrocorax auritus*; and great blue herons, *Ardea herodias*). The PFCA, PFSA, and stable isotope ( $\delta^{15}\text{N}$  and  $\delta^{13}\text{C}$ ) data collected from these species from the Pacific coast of Canada, ranging over 20 to 30 years, were used to investigate temporal changes in PFAS coupled to dietary changes. Perfluorooctane sulfonic acid (PFOS), the dominant PFSA compound in all 4 species, increased and subsequently decreased in auklet and cormorant eggs in line with the manufacturing phase-out of PFOS and perfluorooctanoic acid (PFOA), but concentrations continuously increased in petrel eggs and remained largely unchanged in heron eggs. Dominant PFCA compounds varied between the offshore and coastal species, with increases seen in the offshore species and little or variable changes seen in the coastal species. Little temporal change was seen in stable isotope values, indicating that diet alone is not driving observed PFAS concentrations. *Environ Toxicol Chem* 2015;34:1799–1808. © 2015 SETAC

**Keywords:** Perfluoroalkyl carboxylic acids    Perfluoroalkyl sulfonic acids    Seabirds

## INTRODUCTION

Perfluoroalkyl substances (PFAS) such as sulfonates (PFSAs) and carboxylates (PFCAs) are anthropogenic pollutants used in various industrial and consumer products. Such products include water and stain proofing agents, grease- and oil-resistant packaging for food products, nonstick cookware [1], surfactants, agrochemicals [2], and fire-fighting foams [3], among other uses [4]. Some PFAS are still considered emerging contaminants despite their production since the 1950s [4]. Bioaccumulation potential of PFCAs increases with increasing chain length, with compounds comprising more than 7 fluorinated carbons considered bioaccumulative [5]. The PFSAs, especially longer-chain compounds, are bioaccumulative and biomagnify in the food web [6]. These chemicals are partly hydrophilic and partly lipophilic because of the presence of a carboxylate or sulfonate functional group on the carbon chains [5] and tend to accumulate in protein-rich tissues such as blood and liver [5]. Because of the strength of the carbon-fluorine bond [7], these chemicals are highly resistant to degradation [8], making PFAS a persistent environmental contaminant.

Atmospheric and oceanic water transport can contribute to the long-range dispersal of PFAS [4,9]. Those two routes are likely transport mechanisms for PFAS to remote regions such as the Arctic [10]. Air samples from northwestern Europe indicate

that PFSAs may be directly transported atmospherically on particulates [11], whereas atmospheric deposition has been suggested as a major contributor of PFAS in some regions (e.g., the Baltic Sea) [12]. In part because of that potential for long-range transport, PFAS have been found globally in the terrestrial, marine, and freshwater environments and are now virtually ubiquitous in matrices from those locations, being found in birds [13–16], aquatic organisms [17–19], and mammals [20–23], including humans [19,24]. The deep ocean has been identified as an environmental sink for these chemicals [4,9], possibly via the sedimentation of sinking particles [10]. Thus, a need exists for ongoing monitoring of PFAS across a range of environments. Aquatic birds are ideally situated, in terms of both trophic levels and geographical location, to sample the marine, estuarine, and freshwater environments, and they are among the most widely used indicator species for surveillance work [25].

The long-range transport potential and properties of PFAS have caused concern about possible environmental and toxicological effects, especially neurobehavioral consequences, in animals exposed during early development (reviewed in Houde et al. [10]), and altered inflammatory, adaptive, and immune responses in various animal models (reviewed in Elliott and Elliott [25]). As such, voluntary and regulatory restrictions on use, production, and manufacture of various PFAS have been, and are still being, implemented. In 2000, 3M, the major global producer of PFAS, phased out production of perfluorooctane sulfonic acid (PFOS), perfluorooctanoic acid (PFOA), and related products [26]. Multiple jurisdictions have restricted PFOS and PFOA throughout the 2000s, including the US Environmental Protection Agency (USEPA) [27], Canadian

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environmental and health authorities [28], and the European Union [29]. In addition, PFOS has been included as 1 of the 9 new persistent organic pollutants (POPs) under the Stockholm Convention on POPs since 2009.

Despite those measures, PFAS continue to be detected in the environment [14,20,21,30–32]. Diet is a key exposure route [19,33]; thus, examination of dietary changes using N and C stable isotopes ( $\delta^{15}\text{N}$  and  $\delta^{13}\text{C}$ ) as proxies for trophic position and spatial patterns can help elucidate dietary effects on contaminant concentrations [34]. Bird eggs are widely used for environmental contaminant monitoring, and trophic position can be examined in these same matrices by stable isotope analysis [35,36] to account for variation in contaminant concentrations resulting from dietary changes rather than variation in environmental abundance of contaminants [37].

In light of the national and international phase-outs and restrictions implemented for PFAS over the last 10 to 15 years, and their potential to cause environmental and toxicological harm, in the present study we used long-term data collections to: 1) investigate temporal changes in 13 PFAS in 2 oceanic seabird species (rhinoceros auklets, *Cerorhinca monocerata*, 1990–2010; Leach's storm-petrels, *Oceanodroma leucorhoa*, 1990/1991–2011); and in 2 coastal bird species (double-crested cormorants, *Phalacrocorax auritus*, 1973–2011; great blue herons, *Ardea herodias*, 1982–2012); 2) investigate whether the phase-out of PFOS and PFOA in 2000 has had a noticeable effect on the concentrations of these compounds in these species; and 3) determine whether any relationship exists between PFAS and diet in these populations. In addition, differences between these 4 species and ancient murrelets (*Synthliboramphus antiquus*) were investigated for the most recent year of sampling available, because data for this species were available for 2009.

## METHODS

### Study species

The ancient murrelet (hereafter murrelet) is an offshore, subsurface feeder that preys on zooplankton and small, schooling fish [38]. Adults feed almost exclusively offshore when not on land breeding [38]. Leach's storm-petrel (hereafter petrel) is found throughout the northern Atlantic and Pacific Oceans [39]. It is a planktivorous surface feeder that may feed many hundreds of kilometers beyond the continental shelf edge outside of the breeding season [39]. The rhinoceros auklet (hereafter auklet) is an epipelagic piscivorous feeder [40] inhabiting temperate waters of the northern Pacific [41]. Auklets migrate south in winter [42]. These species return to land to lay a single (auklets, petrels) or 2 eggs (murrelets) annually in a colonial burrow-nesting environment [43,44]. Thus, during the breeding season, including the prelaying period when nutrients are deposited into the egg, auklets feed close (<50 km) to the North American coast, whereas petrels feed up to hundreds of kilometers beyond the North American coast. Little is known about activity during the nonbreeding season, but band recoveries in auklets demonstrate that at least some remain close to the North American coast, whereas petrels roam across most of the North Pacific [45,46]. Double-crested cormorants (hereafter cormorant) and great blue herons (hereafter heron) are both widely distributed across North America [47]. Cormorants are found from resident coastal near-shore species in our Salish Sea study area, western Pacific coast of Canada [47], and herons are year-round residents in freshwater and marine habitats within this region and are a widely used indicator species [48,49].

### Sites, sampling matrix, and design

All eggs were sampled from the Pacific coast of British Columbia, Canada. Murrelet eggs were sampled from Langara Island, and auklet and petrel eggs were sampled from 3 islands—Cleland Island (auklet, petrel), Lucy Island (auklet), and Hippa Island (petrel; Figure 1). All of these islands are distant from large urban centers and located in open coastal positions; thus, direct anthropogenic input is minimal. Auklet and petrel eggs were collected every 4 years during spring and early summer (late April to early July); murrelet eggs were collected in May 2009. Not all sites were sampled in the same years because of cost and logistics (Table 1). Cormorant eggs have been collected from Mandarte Island since 1979, an uninhabited island approximately 60 km southwest of Vancouver (Figure 1). Birds from this colony forage in the South Salish Sea [50]. Heron eggs have been collected from a colony formerly on the grounds of the University of British Columbia and now located in Stanley Park, an urban park of approximately 400 ha located in downtown Vancouver. Herons at both the University of British Columbia and Stanley Park forage in the Fraser River estuary and Burrard Inlet and are therefore referred to throughout as the Fraser River colony. Details on sampling matrix and design have been presented previously [51,52].

Briefly, 15 fresh eggs were collected from individual nests every 4 years for auklets and petrels and stored frozen until prepared for analysis. To conserve archived samples, eggs were analyzed retrospectively as 1 pool of 15 eggs from 1990 to 1991 until either 2002 to 2003 or 2006 to 2007. Eggs were subsequently analyzed as 5 pools of 3 eggs each, as well as being reanalyzed as 1 group of 15, as per previous years. Total number of eggs collected for herons and cormorants has varied over the years, as has year of sampling (Table 1). For each egg, the entire contents were homogenized. Approximately 1 g homogenized content from each egg pool was subsampled and sent for chemical analysis. Subsamples were archived individually and as equal weight pools at  $-40^{\circ}\text{C}$  at the Canadian Wildlife Service National Wildlife Specimen Bank [53]. Moisture and lipid content were recorded for pooled samples.

### Chemical analysis

Samples were analyzed at the National Wildlife Research Centre, Ontario, Canada. Egg contents were homogenized and stored at  $-20^{\circ}\text{C}$  before extraction. The PFAS extraction, cleanup, and analysis for the egg homogenates has been described [13]. Briefly, aliquots of 0.5 g to 1 g wet weight were spiked with labeled internal standards (Supplemental Data, Table S1), and subsequently extracted, cleaned, and fractionated using weak anion exchange solid-phase extraction cartridges. Separation of the target compounds was carried out on Waters model 2695 high-performance liquid chromatography equipped with an ACE 3 C18 analytical column (50 mm  $\times$  2.1 mm D, 3- $\mu\text{m}$  particle size; Advanced Chromatography Technologies) coupled to a Waters Quattro Ultima triple quadrupole mass spectrometer (Waters). An electrospray ionization source in negative mode was used for analysis of PFCAs and PFSAs. Quantification was performed by using an internal standard approach.

### Quality control

A known amount of labeled compound (surrogate) was added to every sample before extraction because the native and labeled analogs exhibit similar effects on extractions,

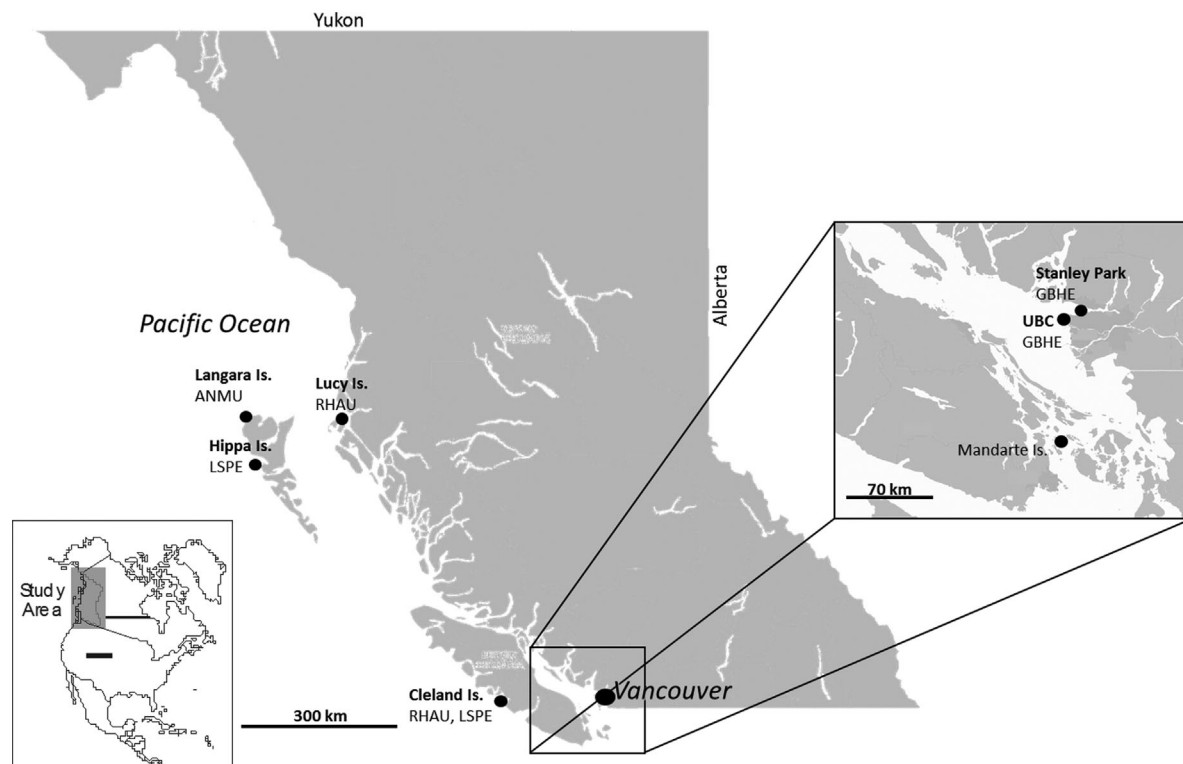


Figure 1. Map of the Pacific coast of British Columbia, Canada, indicating the approximate location of each sampled seabird colony. Inset, bottom left: Gray area indicates the Pacific coast of Canada in relation to the North American continent. Inset, right: Sampled colonies of great blue heron (GBHE; *Ardea herodias*) and double-crested cormorant (*Phalacrocorax auritus*). RHAU = rhinoceros auklet (*Cerorhinca monocerata*); LSPE = Leach's storm petrel (*Oceanodroma leucorhoa*); and ANMU = ancient murrelet (*Synthliboramphus antiquus*).

concentration, and liquid chromatography. By using the surrogate responses from the sample run and the relative response factor values, the concentrations of PFCA and PFSA were recovery corrected. Method limit of detection was determined using in-house validation methods with an equivalent 3:1 signal-to-noise ratio. The limit of detection for all examined PFAS was 0.1 ng/g, with the exception of perfluorodecane sulfonic acid (PFDS), which was 0.2 ng/g. To check for contamination of analytes from solvents or the extraction process, 1 blank sample was prepared and analyzed with every batch of egg pools ( $n = 10$ ). An in-house reference material of double-crested cormorant egg homogenate (based

on eggs collected in 2003) was analyzed to assess reproducibility of the method. Recovery efficiencies of the PFAS internal standards were generally greater than 60%.

#### Stable isotope analysis

Stable isotopes of delta 13 carbon ( $\delta^{13}\text{C}$ ) and delta 15 nitrogen ( $\delta^{15}\text{N}$ ) were analyzed from the same egg homogenates as the chemical analyses. Detailed methods have been reported previously [36,51]. Subsamples of egg homogenate were prepared and sent to the Stable Isotope Facility, University of California, Davis, for analysis. All  $\delta^{13}\text{C}$  values were lipid normalized, because differences in lipid content can obscure

Table 1. Seabird species and years sampled at each site on the Pacific coast of Canada<sup>a</sup>

Species	Site	Coordinates	Years Sampled
Ancient murrelet	North coast, Langara Island	54°12'N, 133°1'W	2009
Rhinoceros auklet	South coast, Cleland Island	49°10'N, 126°5'W	1990, 1994, 1998, 2002, 2006, 2010
	North coast, Lucy Island	54°18'N, 130°37'W	1990, 1995, 1999, 2003, 2006, 2010
Leach's storm petrel	South coast, Cleland Island	49°10'N, 126°5'W	1990, 1994, 1998, 2002, 2006, 2011
	North coast, Hippa Island	53°26'N, 132°59'W	1991, 1995, 1999, 2003, 2007, 2011
Double-crested cormorant	South coast, Mandarte Island	43°38'N, 123°17'W	1973, 1985 (PFASs only), 1990 (PFASs only), 1994, 1998, 2002, 2006, 2011
Great blue heron	South coast, Fraser River	49°30'N, 123°14'W 49°26'N, 123°25'W	1982 (PFASs only), 1987 (PFASs only), 1992, 1996, 2000, 2003, 2004, 2008, 2012

<sup>a</sup>Sampling occurred between April and July (from Leat et al. [64]). PFASs = perfluoroalkyl sulfonates.

variation in  $\delta^{13}\text{C}$  [36]. However, the Suess effect on  $\delta^{13}\text{C}$  [54] was not corrected for in the present study, given the relatively short time span of the data set presented ( $\sim 20\text{--}30$  yr) compared with the timing of the Suess effect.

#### Statistical analysis

Where relevant, concentrations are given on a nanogram per gram wet weight basis, unless otherwise specified. Compounds that were below the limit of detection were divided by 2 to give a conservative estimate of concentration. Arithmetic means were calculated for each year in which multiple pooled samples were available for species and site. Where applicable,  $\Sigma\text{PFSA}$ s include perfluorobutane sulfonic acid (PFBS), perfluorohexane sulfonic acid (PFHxS), PFOS, and PFDS, whereas  $\Sigma\text{PFCA}$ s include perfluorohexanoic acid (PFHxA), perfluoroheptanoic acid (PFHpA), PFOA, perfluorononanoic acid (PFNA), perfluorodecanoic acid (PFDA), perfluoroundecanoic acid (PFUdA), perfluorododecanoic acid (PFDoA), perfluorotridecanoic acid (PFTTrDA), and perfluorotetradecanoic acid (PFTeDA). Any temporal analysis excludes murrelets, because only data from 2009 were available for this species.

All dominant compounds were log transformed before running either simple linear regression or quadratic regression to examine temporal changes. Doubling and halving times of log-transformed dominant compounds for each species before 2000 and after 2000 were conducted using the slope of a simple regression line and assuming first-order (exponential) increases where appropriate. This year was chosen because 3M phased out PFOS, PFOA, and related compounds from 2000 to 2002 [26]. In all cases  $n = 3$ , except before 2000 for cormorants, where  $n = 5$ ; hence, results should be interpreted with caution.

A principal component analysis was used to examine the pattern of distribution of log transformed dominant compounds and other quantifiable compounds over the entire time period between all 4 species ( $n = 6$  for each species and site combination for auklets, petrels, and cormorants;  $n = 7$  for herons). General linear models were used to examine the relationship between  $(\log)\Sigma\text{PFSA}$  concentrations and  $(\log)\Sigma\text{PFCA}$  concentrations, with species (auklet, petrel, cormorant, and heron), site (offshore and coastal), and lipid percentage as independent variables, and included both linear and quadratic terms for year to account for nonlinearity in the relationship with year. Because species and site had a significant effect on both  $(\log)\Sigma\text{PFSA}$  and  $(\log)\Sigma\text{PFCA}$  concentrations over time, data were further divided into offshore (auklet and petrel) and coastal (cormorant and heron) species and reexamined. General linear models were run for  $(\log)\Sigma\text{PFSA}$  and  $(\log)\Sigma\text{PFCA}$  concentrations in auklets and petrels using lipid percentage, site, species, year, and year<sup>2</sup>, and repeated for cormorants and herons.

Simple linear regression was used to examine temporal trends of stable isotopes of  $\delta^{13}\text{C}$  and  $\delta^{15}\text{N}$ . One outlier (petrel, Hippa Island,  $\delta^{15}\text{N}$ , 0.32%;  $\delta^{13}\text{C}$ ,  $-29.57\%$ ) was removed from the data set. Multiple linear regression was used to examine whether any relationships existed between  $\delta^{13}\text{C}$ ,  $\delta^{15}\text{N}$ , and lipid percentage to log natural transformed  $\Sigma\text{PFSA}$  and  $\Sigma\text{PFCA}$  concentrations from all species collectively, and by species and site over time. Statistical analyses were conducted in Excel 2010 and R Ver 3.0.3. Significance level for all tests performed was set to  $p < 0.05$ .

## RESULTS AND DISCUSSION

#### Dominant compounds

The dominant PFSA compound for all species examined was PFOS ( $\text{C}_8$ ). In murrelet eggs sampled in 2009, PFOS constituted

more than 95% of  $\Sigma\text{PFSA}$ s; in every sampling year,  $\Sigma\text{PFSA}$ s constituted more than 97% in petrel eggs, more than 89% in auklet eggs, and more than 78% in cormorant eggs; and in heron eggs, PFOS constituted more than 60% of  $\Sigma\text{PFSA}$ s in every sampling year (Supplemental Data, Table S2). Perfluorooctane sulfonic acid is typically the most commonly detected PFSA in wildlife [5,10,30] regardless of species, tissue, or geographical location [20–23,55]. Concentrations observed are within the range of concentrations seen in other seabird species from, for example, Nunavut (Canada; 6.5–37 ng/g wet wt) [30], the Laurentian Great Lakes (83–933 ng/g wet wt) [31], and the North Sea (Germany; 20–170 ng/g wet wt) [56], but considerably lower than the maximum concentration observed from murre (*Uria aalge*) eggs in the Baltic Sea (25–1324 ng/g wet wt) [32]. Perfluorodecane sulfonic acid ( $\text{C}_{10}$ ) also showed quantifiable levels in all years for auklet eggs at both sites, although concentrations were far below that of PFOS, whereas PFDS was below 0.5 ng/g wet weight in murrelet eggs and remained below the limit of detection in petrel eggs except in 2011 at the south coast site. In the 2 coastal species, PFDS showed a reasonable contribution to  $\Sigma\text{PFSA}$ s, in particular in herons, in which it contributed up to 37% to  $\Sigma\text{PFSA}$ s in some years (Supplemental Data, Table S2). Similarly, PFDS was detected at low concentrations in herring gull (*Larus argentatus*) eggs collected from colonies throughout the Laurentian Great Lakes [31].

The dominant PFCA compounds for all 3 offshore species at all sites were 2 long-chained odd-numbered PFCAs, PFUdA ( $\text{C}_{11}$ ), and PFTTrDA ( $\text{C}_{13}$ ). This pattern of long-chained odd-numbered PFCAs being detected at higher concentrations compared with even-numbered long-chained PFCAs has been reported in various species, such as beluga whales (*Delphinapterus leucas*) from Alaska [23], various Canadian Arctic biota (e.g., polar bears, *Ursus maritimus*; mink, *Neovison vison*; arctic fox, *Vulpes lagopus*; ringed seals, *Phoca hispida*; common loons, *Gavia immer*, and so forth) [57], herring gull eggs from Norway [58], and seabird species from the Canadian Arctic [30]. Dominant PFCA compounds in coastal species differed somewhat from the offshore species. In cormorants, PFOA ( $\text{C}_8$ ) and PFNA ( $\text{C}_9$ ) were the major contaminants. In herons, PFNA was initially dominant (1992, 1996), followed by PFTTrDA; however, PFTTrDA surpassed PFNA in all subsequent years, as did PFDoA ( $\text{C}_{12}$ ) and PFUdA (Supplemental Data, Table S2). Other PFCA compounds found in quantifiable concentrations in the offshore species were PFNA, PFDA, PFDoA, and PFTeDA, and in coastal species PFHxA and PFHpA. All other compounds were either not found or were below the limit of detection.

#### Temporal trends

Perfluorooctane sulfonic acid contributed the most to total concentrations of PFAS in both offshore and coastal species regardless of location, although the total contribution was decreasing over time, whereas in the offshore auklets and petrels, total contribution of PFTTrDA and PFUdA in particular increased with time. Perfluorooctane sulfonic acid concentrations increased in auklet eggs at both sites (Figure 2A, 2B) and in cormorant eggs (Figure 2E) until the early 2000s, when concentrations decreased. The PFOS contribution to total PFAS generally has decreased since the late 1990s (cormorants) and early 2000s (auklets; Table 2), in line with industry phase-outs [26]. Halving times for PFOS in auklet eggs after 2000 were  $-12.7$  years and  $-21$  years (south and north coast, respectively), whereas doubling time was 96.3 years for

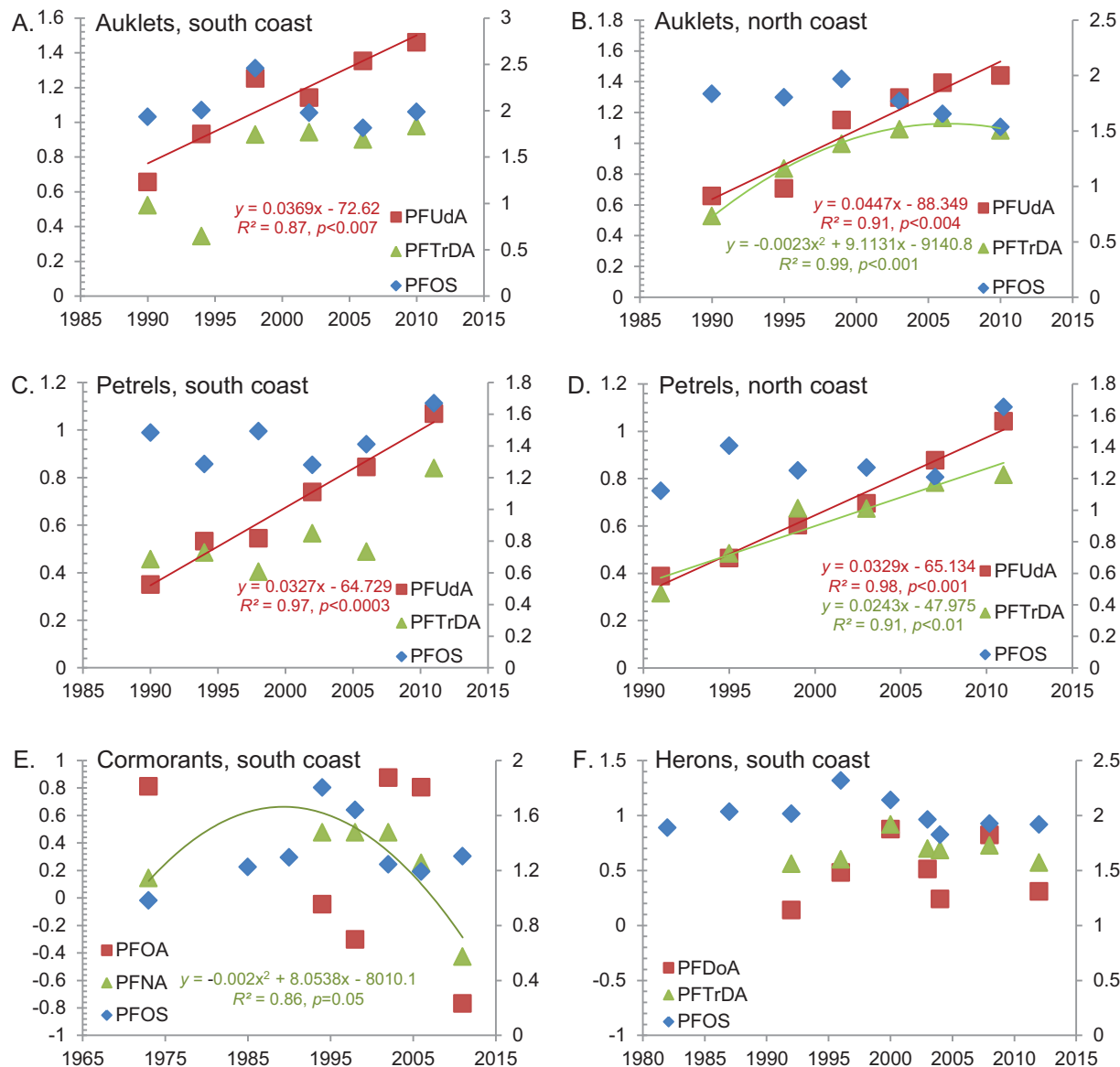


Figure 2. Log(10) transformed perfluoroundecanoic acid (PFUdA) and perfluorotridecanoic acid (PFTrDA; primary y axis) and perfluorooctane sulfonic acid (PFOS; secondary y axis), over time for (A) rhinoceros auklets from the south coast; (B) rhinoceros auklets from the north coast; (C) Leach's storm-petrels from the south coast; and (D) Leach's storm-petrels from the north coast; (E) log(10) transformed perfluorooctanoic acid (PFOA) and perfluorononanoic acid (PFNA; primary y axis) and PFOS (secondary y axis) for double-crested cormorants from the south coast; and (F) log(10) transformed PFTrDA and perfluorododecanoic acid (PFDoA; primary y axis) and PFOS (secondary y axis) for great blue herons from the Fraser Estuary. Regression line and equation are only shown where a significant temporal change was observed.

cormorant eggs (Table 2). Comparably, an increasing trend for PFOS was observed in murre eggs from the Baltic Sea between 1968 (25 ng/g wet wt) and 2005 (614 ng/g wet wt) [32]; however, more recently (2001–2011) a decreasing trend has been indicated in this data set [59]. Perfluorooctane sulfonic acid concentrations in peregrine falcon (*Falco peregrinus*) eggs sampled from Sweden between 1974 and 2007 increased rapidly during the 1970s and early 1980s before leveling off [14].

By contrast, PFOS concentrations increased over the entire period in petrel eggs at both sites (Figure 2C, 2D), whereas in heron eggs PFOS concentrations showed little change over time (Figure 2F). The PFOS doubling times after 2000 in petrels were 15.9 years and 14.4 years (south and north coast, respectively), considerably shorter compared with times before 2000 of 693 years and 43 years, respectively (Table 2), indicating that concentrations of these compounds continue to increase in the oceanic environment and that phase-outs have not had a

noticeable effect on contamination of petrels at those locations. By contrast, PFOS doubling time before 2000 in herons was 40.1 years and 385.1 years after 2000, indicating slowing accumulation in this species. Alternatively, because petrels spend the nonbreeding season in offshore pelagic environments, potentially closer to Asia, they may be less susceptible to regulatory actions in North America and may be affected by ongoing use in Asia. Conversely, PFOS in murre (*U. lomvia*) and fulmar (*Fulmarus glacialis*) eggs sampled from Prince Leopold Island in the Canadian Arctic between 1975 and 2011 did not change significantly over this period [30], also indicative of minimal effects wrought by phase-outs and other more recent North American and European regulations.

Despite regulations and restrictions on production and use in Europe [29] and North America [27,28], China began large-scale production of PFOS in 2003 and has continued to produce PFOS for use in various industries [60–62]. Most PFAS

Table 2. Doubling and halving times (in yr) for pre- and post-2000 values from simple linear regression for log transformed dominant compounds in rhinoceros auklet, Leach's storm petrel, double-crested cormorant, and great blue heron eggs<sup>a</sup>

Compound	Rhinoceros auklets, south coast		Rhinoceros auklets, north coast	
	Before 2000, <i>n</i> = 3	After 2000, <i>n</i> = 3	Before 2000, <i>n</i> = 3	After 2000, <i>n</i> = 3
PFOS	10.6	630.1	49.5	(21.0)
PFUdA	9.3	18.8	13.1	34.5
PFTTrDA	13.6	39.6	13.3	(301.4)
Compound	Leach's storm-petrels, south coast		Leach's storm petrels, north coast	
	Before 2000, <i>n</i> = 3	After 2000, <i>n</i> = 3	Before 2000, <i>n</i> = 3	After 2000, <i>n</i> = 3
PFOS	693.2	15.9	42.8	14.4
PFUdA	28.5	18.7	25.9	16.0
PFTTrDA	(106.6)	21.7	15.5	39.2
Compound	Double-crested cormorant, south coast		Great blue herons, south coast	
	Before 2000, <i>n</i> = 5 (PFSA), <i>n</i> = 3 (PFCA)	After 2000, <i>n</i> = 3	Before 2000, <i>n</i> = 4	After 2000, <i>n</i> = 4
PFOS	22.7	96.3	40.1	385.1
PFDS	0.0	(26.3)	Quadratic	(10.4)
PFOA	(16.0)	(3.7)	3.5	Quadratic
PFNA	49.2	(6.8)	(43.3)	7.5
PFUdA	11.2	4.5	22.8	6.3
PFTTrDA	9.8	15.8	16.5	(59.8)

<sup>a</sup>Halving times are shown in parentheses for regressions that showed declining trends.

PFSA = perfluoroalkyl sulfonate; PFCA = perfluoroalkyl carboxylate; PFOA = perfluorooctanoic acid; PFNA = perfluorononanoic acid; PFUdA = perfluoroundecanoic acid; PFTTrDA = perfluorotridecanoic acid; PFOS = perfluorooctane sulfonic acid; PFDS = perfluorodecane sulfonic acid.

producers are located in coastal China [63]. Increased production in China may have offset any progress in reducing PFOS gained by other industry phase-outs with respect to exposure of some Arctic biota [30]. Although little is known about their nonbreeding ground movements, petrels may spend some portion of the nonbreeding season in the western Pacific, likely in tropical waters [45]. Overwintering exposure has been shown to have a significant effect on polybrominated diphenyl ether (PBDE) concentrations in great skua (*Stercorarius skua*) [64]. The elimination half-life of PFOS is rather variable, ranging from an estimated 13.6 days in mallard (*Anas platyrhynchos*) serum and 20.1 days in northern bobwhite quail (*Colinus virginianus*) serum [65] to 125 days in male chicken serum (*Gallus gallus*) [66], more than 89 d in male rat serum [65], and 100 days to 200 days in cynomolgus monkey (*Macaca fascicularis*) serum [67], to 8.6 years in humans [68]. Thus, the continued increasing PFOS trends observed in petrels may be attributable to carryover from winter exposure. Furthermore, if breeding-ground exposure was solely accountable for the observed concentrations, expecting similar temporal trends in auklet and petrel eggs from the south coast, closer to industrial regions in southern Canada and the United States, would be reasonable.

For both offshore species at all sites,  $\Sigma$ PFCA showed increasing temporal trends. Perfluoroundecanoic acid showed consistent, and mostly significant, increasing temporal trends in both species at all sites (Figure 2A–D). Perfluorotridecanoic acid showed significant increasing trends in auklet eggs at Cleland Island (Figure 2A), petrel eggs at Hippa Island (Figure 2D), and general increasing trends in petrel eggs from Cleland Island (Figure 2C). However, in auklet eggs from Lucy Island, an increase in PFTTrDA concentration was followed by a significant decrease (Figure 2B). Significant increases

( $p < 0.01$ ) were found in  $\Sigma$ PFCA for both auklets and petrels at all sites (not shown). By contrast,  $\Sigma$ PFCA in the coastal species showed some decrease in concentration in the most recent years (not shown). This pattern was also observed in the dominant PFCA compounds (Figure 2E, 2F), which is most likely a reflection of the proximity to urban sources.

Similar to our findings, PFUdA and PFTTrDA were found to significantly increase in eggs of thick-billed murres and northern fulmars from Prince Leopold Island, Canadian Arctic, between 1975 and 2011 [30]. At 3 sites in the North and Baltic Sea, longer-chained PFCA increased over the last 20 yr in herring gull eggs [56]. The  $\Sigma$ PFCA concentrations increased significantly in the eggs of 4 of 7 herring gull colonies sampled from the Laurentian Great Lakes in 1990 and 1997 to 2010 [31]; in herring gull eggs from 2 colonies in northern Norway from 1983 to 1993, and then showed either a weak increase after 1993 or a leveling off [58]. Both PFUdA and PFTTrDA are unintentionally produced substances [69]. Instead, their presence in the environment is probably attributable to both direct (impurities in PFOA and PFNA production) and indirect sources (atmospheric transport and degradation of precursors such as the ammonium salt of PFNA [69]).

Although an agreement was made in the United States in the mid-2000s between the USEPA and 8 major global companies to decrease emissions and product content of PFOA, higher homologues, and precursors by 95% by 2010 [27,69], and in Canada an agreement was made in 2010 between Environment Canada, Health Canada, and 4 companies from the perfluorinated product industry to restrict PFCA in products [28], clearly not enough time has elapsed to observe a related effect on  $\Sigma$ PFCA concentrations in petrel and auklet eggs. However, a general decrease in recent sampling years was seen in the coastal species. Further

monitoring is warranted to track PFCA development in light of North American agreements.

### Species differences

Differences are apparent, especially between the coastal and offshore species. In the most recent year of sampling, PFOS concentration in herons from a colony located within the city of Vancouver that feed in the Fraser River estuary were the greatest among the sampled species. Surprisingly, PFOS concentration was second lowest in cormorants, only being lower in ancient murrelets, despite the relative proximity of the cormorant colony to urbanized areas in the Salish Sea (Table 1, Figure 3A). Concentrations of PFUdA and PFTrDA in the 2 coastal species were considerably lower compared with the offshore species (Figure 3B), indicating a difference in exposure sources between coastal and offshore; for example, coastal species are exposed because of their relative proximity to major cities, and where regulations on PFAS appear to be having a positive effect. It appears that the more offshore species may be exposed during nonbreeding ground exposure or via redistribution of these compounds to the pelagic environment from urban sources.

Little variation was seen in spatial pattern of the 3 dominant compounds between the offshore species. Those results support the idea that PFAS levels in eggs of the offshore species primarily reflect exposure during the 7 months/year to 8 months/year both species spend on nonbreeding grounds rather than the 4 to 5 months/year that they spend at the breeding grounds [44], especially because nutrients in the eggs are deposited at the start of the breeding season, before they have several months' recent exposure. We suggest either that egg nutrients (including PFAS) are deposited at varying times after arriving from the nonbreeding grounds or that they are derived from tissues that have a longer PFAS half-life than the 14 to 20 days reported for some avian species [65] and may be similar to that seen in chicken serum [66]. However, clear differences are seen among the species; herons had the highest PFOS concentration, cormorants had the lowest concentrations of both dominant PFCAs, and the offshore species had the highest concentrations of dominant PFCAs. Those observations are supported by general linear model results examined for the entire time series for each species (ancient murrelet excluded), where species and site had a significant effect on log transformed  $\Sigma$ PFSA and  $\Sigma$ PFCA concentrations when all 4 species were combined

( $p < 0.0001$ ,  $p < 0.0001$  for site and species for both  $[\log] \Sigma$ PFSA and  $[\log] \Sigma$ PFCA, respectively), as did lipid percentage on  $[\log] \Sigma$ PFSA ( $p < 0.04$ ) but not  $(\log) \Sigma$ PFCA. Species differences could be attributable to differing exposure, proximity to urban centers and sources, and dietary and possibly metabolism differences.

More than 70% of the variance was explained by the first 2 axes of a principal component analysis examining the pattern of distribution of log-transformed dominant PFAS compounds and other quantifiable compounds among the 4 species (murrelets excluded) over the whole sampling period (Figure 4). Minimal overlap was seen between auklets, herons, and cormorants, and no overlap was seen with petrels. Petrels appear to be uninfluenced by any particular compound or group of compounds over time. In contrast, the coastal species appear to be more influenced by PFSA compounds, and auklets are more influenced by PFCA compounds, again indicating species differences in PFAS exposure. No clear shift in compound influence was seen before 2000 or after 2000 for any species.

### Stable isotope analysis

Neither C nor N stable isotope ratios varied much over time from auklet or petrel eggs at any site, and murrelet egg stable isotope values were reasonably stable among individuals for the single year sampled. For cormorants,  $\delta^{15}\text{N}$  varied little over time, whereas  $\delta^{13}\text{C}$  showed a nonsignificant increase. Based on earlier dietary assessments, auklets and petrels are offshore pelagic feeders [70], as indicated by  $\delta^{13}\text{C}$  values, with petrels being the more pelagic of the 2, as expected. Cormorant egg  $\delta^{13}\text{C}$  values ( $-13.2\text{‰}$  to  $-17.6\text{‰}$ ) are more enriched compared with auklet ( $-17.3\text{‰}$  to  $-19.4\text{‰}$ ), murrelet ( $-18.5\text{‰}$  to  $-19.7\text{‰}$ ), and petrel eggs ( $-20.7\text{‰}$  to  $-23.7\text{‰}$ ) indicating that auklets, murrelets, and petrels feed more offshore compared with cormorants, with petrels feeding the farthest from shore. Assuming that a fractionation factor of 3.2‰ to 3.4‰ represents an average for multiple trophic transfers [34], neither auklet nor petrels transcended more than 1 trophic level over the examined period. However, cormorant  $\delta^{15}\text{N}$  values spanned a range of 3.7‰, and  $\delta^{13}\text{C}$  values spanned a range of 4.4‰, indicative more of temporal dietary variation rather than a change in trophic feeding level given the nonsignificant changes seen over time for both of these stable isotopes. Values for  $\delta^{15}\text{N}$  were between 11‰ and 16‰ for auklets, murrelets, and petrels, and 13‰ and 17.5‰ for cormorants. When temporal

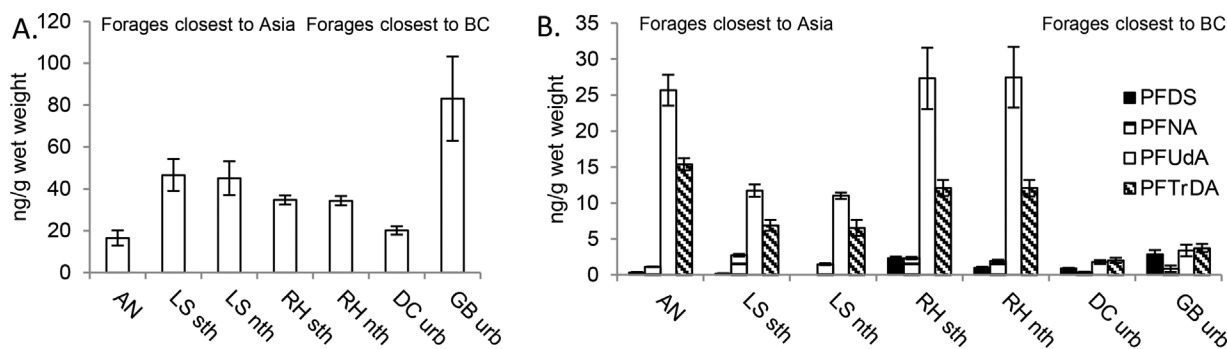


Figure 3. (A) Average perfluorooctane sulfonic acid (PFOS) concentration (ng/g wet wt  $\pm$  standard error of the mean [SEM]) and (B) average perfluorodecane sulfonic acid (PFDS), perfluorononanoic acid (PFNA), perfluoroundecanoic acid (PFUdA), and perfluorotridecanoic acid (PFTrDA) concentrations (ng/g wet wt  $\pm$  SEM) in ancient murrelet, rhinoceros auklet, Leach's storm-petrel, double-crested cormorant, and great blue heron eggs for the most recent sampling year. AN = ancient murrelet, Langara Island (north coast; 2009); RH sth = rhinoceros auklets, Cleland Island (south coast; 2010); RH nth = rhinoceros auklets, Lucy Island (north coast; 2010); LS sth = Leach's storm-petrels, Cleland Island (south coast; 2011); LS nth = Leach's storm-petrels, Hippa Island (north coast; 2011); DC urb = double-crested cormorants (south coast, urban; 2011); GB urb = great blue herons (south coast, urban; 2012).



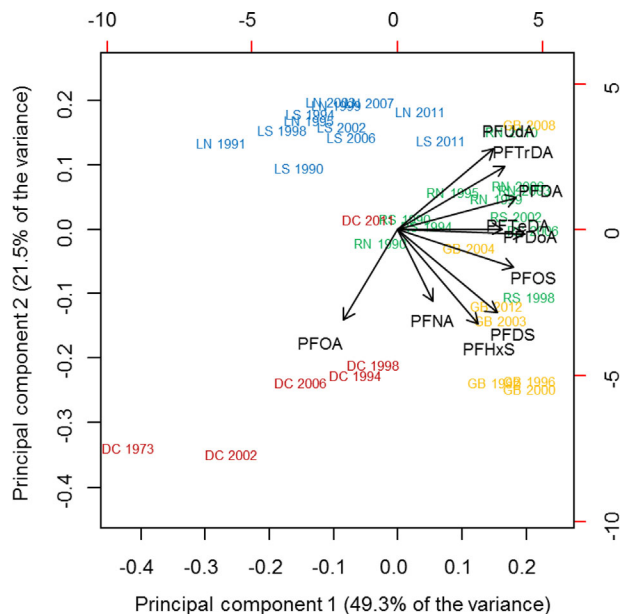


Figure 4. Principal component analysis of all compounds that showed quantifiable concentrations for rhinoceros auklets at the south coast (RS; green), rhinoceros auklets at the north coast (RN; green), Leach's storm-petrels at the south coast (LS; blue), Leach's storm-petrels at the north coast (LN; blue), double-crested cormorants (DC; red), and great blue herons (GB; yellow) for the whole monitoring period. Years are shown next to each species and site; for example, DC 2002 indicates double-crested cormorants, 2002.

stable isotope data for auklet, petrel, and cormorants were combined, no significant relationships were observed between  $\delta^{13}\text{C}$ ,  $\delta^{15}\text{N}$ , or lipid percentage and  $(\log)\Sigma\text{PFSA}$ , and  $\delta^{13}\text{C}$  and  $(\log)\Sigma\text{PFCA}$ . The lack of relationships and variation in  $\delta^{13}\text{C}$  and  $\delta^{15}\text{N}$  over time indicates that diet has not changed in any marked way for these bird species. Therefore, diet alone is unlikely to be driving the variation in observed concentrations in PFAS in these species.

#### Toxicological implications

Various effects have been observed from laboratory experiments examining PFOS exposure in birds, ranging from decreased pipping ability from doses as low as  $0.1\ \mu\text{g}\ \text{PFOS/g}$  egg weight [71–73], to pathological changes in the liver, brain asymmetry, and increases in spleen mass and plasma lysozyme activity indicative of innate immune function from doses of  $1\ \mu\text{g/g}$  to  $5\ \mu\text{g/g}$  egg weight [73,74], to reductions in body weight [75]. O'Brien et al. [71] examined PFUdA and PFDS in chicken eggs but found no effect on pipping success at concentrations up to  $10\ \mu\text{g/g}$  egg weight. Estimated median lethal dose values range from  $4.9\ \mu\text{g/g}$  egg weight to  $93\ \mu\text{g/g}$  egg weight [72,73], although confidence intervals vary over several orders of magnitude. A toxic reference value of  $1.7\ \mu\text{g/g}$  egg has been reported for northern bobwhite quail and mallard duck eggs [75,76] (assuming 1 mL egg corresponds to 1 g). The lowest-observable-adverse-effect level (LOAEL) for PFOS has been estimated as  $0.1\ \mu\text{g}\ \text{PFOS/g}$  egg weight [73]. Maximum PFOS concentrations observed in herons and auklets from the south coast colony of Cleland Island were above the LOAEL for PFOS, where potential effects on pipping success may be observed. However, the most recent year of sampling shows PFOS concentrations in all 4 species to be below the LOAEL. Further monitoring and enforcement of agreements and regulations implemented are necessary, and more research on toxicological effects of exposure to PFCA compounds is

needed. If concentrations of some of these PFAS compounds continue to increase, further toxicological data, other than single-species egg injection studies, on exposure to birds may be warranted.

#### SUPPLEMENTAL DATA

##### Tables S1–S2. (78 KB DOC).

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**Data availability**—Yearly averages for contaminants examined are available in the present study as supplemental data. Further details can be requested from the authors (John.Elliott@ec.gc.ca).

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